SEROPREVALENCE OF EQUINE HERPES VIRUS TYPE-1 IN HORSES AND DONKEYS IN QALUBIAH GOVERNORATE IN EGYPT

Hazem M. El Moghazy¹, Mohamed G. Abdelwahab², Faiysal I. Hamouda², Elsayed M. Ibrahim² and Safaa M.A. Warda³

¹ Veterinary Teaching Hospital, Faculty of Veterinary Medicine, Benha University.
 ² Animal Medicine Department, Faculty of Veterinary Medicine, Benha University.
 ³ Veterinary Serum and Vaccine Research Institute, Abbasia, Cairo, Egypt.

ABSTRACT

Equine hepesirus-1 (EHV-1) is among the infectious diseases that threaten equine health. EHV-1 infects horses causing epidemic respiratory disease, abortion in pregnant mares, neonatal foal death, myeloencephalopathy, and chorioretinpathy. The present study was designed to investigate the presence of EHV-1 in blood serum of horses and donkeys in different localities in Qalubiah Governorate. A total of 185 blood serum samples were collected from horses and donkeys. All animals were evaluated based off whether they were of Indigenous breeds, different ages, apparent health, and clinical signs to equine herpes. The collected blood serum samples were investigated against equine herpes virus type-1 by using an indirect ELISA. The results revealed that 39.5% (73/185) of all horses and donkeys' sera samples were EHV-1 seropositive while 42.86 % (33/77) of the horses' sera samples and 37 % (40/108) of the collected donkeys' sera samples were EHV-1 seropositive. Meanwhile, 57.14 % (44/77) in horses and 63 % (68/108) in donkeys' sera samples were found to be seronegative for the EHV-1 infections. Analysis of data revealed that, young animals (<2 years) were represent (50%) (14/28) while adult animals $(\geq 2years)$ were (37.6%) (59/157) EHV-1 antibodies seropositive. Further analysis showed that, EHV-1 antibodies seropositive young horses (9/17) (52.9%) were higher than their adults (40%) (24/60). While, EHV-1 antibodies seropositive young donkeys (45.5%) (5/11) were higher than their adults (36.1%) (35/97). In the present study, the total number of (185) examined horses and donkeys' sera were differentiated into (79) males and (106) females. Out of (79) male cases, 15.2% (12/79) of total male horses and donkeys were EHV-1 seropositive while 15.15% (5/33) of male horses' sera and 15.2% (7/46) of male donkeys' sera were EHV-1 seropositive. However, out of (106) female cases, 57.5% (61/106) of total female horses and donkeys were EHV-1 seropositive. While, the percentage of seropositive sera samples of female horses and donkeys were 63.6% (28/44) and 53.2% (33/62) respectively.

Key Words: EHV-1; Seroprevalence; indirect ELISA.

1. INTRODUCTION

Equine herpes virus (EHV) is a highly contagious disease that affects equids probably caused by either of the two closely related herpes viruses, equine herpes virus-1 (EHV-1) or equine herpes virus-4 (EHV- 4) (AAEP, 2013).

Equine herpes virus-1 (EHV-1) is an important equine viral pathogen that exerts its major impact by inducing abortion storms or sporadic abortions in pregnant mares, early neonatal death in foals, respiratory disease in young horses and myeloencephalopathy in adult horses (van Maanen, 2002; Patel and Heldens, 2005).

Infections caused by EHV-1 are particularly common in young horses, and typically result in establishment of latent infection within the 1st weeks or months of life (Foote, et al., 2004) with subsequent viral reactivation causing clinical disease and viral shedding during periods of stress. Therefore, rapid diagnostic methods are useful for managing the disease (OIE, 2015).

Due to latency being a general feature of the herpes viruses, the conduct of routine serological tests bears significance for the control of herpes virus infections (Siedek, et al., 1999). Thus, ELISA is frequently used in the laboratory diagnosis of EHV-1 and EHV-4 as a serological test (Yasunaga, et al., 1998 and Gilkerson, et al., 1999).

The goal of this research was to determine the actual situation of EHV-1 infections in horses and donkeys in Qalubiah Governorate and to make recommendations for testing and dealing with infections in order to prevent economic losses.

2.MATERIAL AND METHODS

2.1. Materials:

2.1.1. Animals:

2.1.1.1. Horses and donkeys:

One hundred and eighty five horses and donkeys of different ages and sexes from different localities in Qalubiah Governorate in addition to the animals conducted to the veterinary Teaching Hospital, Faculty of Veterinary Medicine, Benha University) as shown in Table (1 & 2).

Table (1): Number of horses and donkeys used for seroprevalence of EHV-1 antibodies in Qalubiah
 Governorate, according to the age.

Species	Ani				
Age	Horses	Donkeys	Total No.		
< 2 years	17	11	28		
2-5 years	26	28	54		
>5 years	34	69	103		
Total	77	108	185		

Table (2): Number of horses and donkeys used for seroprevalence of EHV-1 antibodies in Qalubiah
 Governorate, according to the sex.

Sex	Anima		
	Horses	Donkeys	Total No.
Male	33	46	79
Female	44	62	106
Total	77	108	185

2.1.2. Antigen:

A partially purified antigen of EHV-1 local isolate propagated on Vero cell line was prepared according to the method described by **Dutta et al.**, (1983) and Azmi and Field (1993) used in serological test.

2.1.3. Antisera:

Horse anti-EHV-1 hyperimmune sera, prepared in Veterinary Serum and Vaccine Research Institute, Abbasia, Cairo, used as positive control in serological test.

2.1.4. Reagent and solution for serological test:

2.1.4.1. Reagent and solutions used in solid phase ELISA;

2.1.4.1.1. Phosphate buffered saline (PBS):

Sodium chloride	8.0 gm
Potassium chloride	0.2 gm
	0
Potassium dihydrogen phosphate	0.2 gm
Di-sodium hydrogen phosphate 2 H2O	3.1 gm
DI-sourum nyurogen phosphate 2 1120	J.1 gm
Deionized Distellad water to	1000 ml
2.1.4.1.2. Coating buffer:	
Na ₂ CO3	1.86 gm
NaHCO3	2.93 gm
DDW	1000 ml
2.1.4.1.3. Washing buffer:	

PBS + 0.05 % Tween 20

2.1.8.2.4. Blocking buffer:

PBS + 2.3 % bovine serum albumin

2.1.4.1.5. Diluting buffer:

PBS + 0.05 % Tween 20 + 0.1 % bovine serum albumin

2.1.4.1.6. Substrate solution:

2.1.4.1.7. Substrate working so	lution:	-
DDW	1000 ml	pH 5.5-6.0
Sodium dihydrogen phosphate	17.3 gm	
Citric acid	7.74 gm	

This was freshly made by addition of ortho-phenelene diamine-2 HC1 (OPD), Sigma in a concentration of 1 gm per 1 ml of substrate stock solution and 0.8 ul of H_2O_2 (30%) per 1 ml buffer added just before use and kept in a dark brown bottle.

2.1.4.1.8. Anti-horse IgG:

Anti-horse IgG whole molecule conjugated with preoxidase produced by binding site Birmingham Research Park, UK.

2.1.4.1.9. Stopping solution: ' •

2 N sulphuric acid solution prepared by mixing 1 ml concentrated sulphuric acid with 6.5 ml water to be used immediately.

2.2. Methods:

2.2.1. Samples collection from horses:

Volumes of 6 to 8 mL blood samples were collected from each of the Egyptian horses and donkeys, using external jugular venipuncture method. Species, age, sex, previous vaccination and last trip history were also recorded.

2.2.2. Preparation of blood serum samples:

The serum samples were prepared according to the standard procedures after blood clotting at ambient environment and overnight incubation at $18 - 26^{\circ}C$ controlled room temperature. Serum samples were separated to stock tubes and kept in a freezer (-20 °C) until use.

2.2.3. Enzyme linked immunosorbent assay (Solid phase ELISA):

• Partially purified EHV-1 antigen was diluted 1:100 in coating buffer.

• A Microtiter plate (96-wells flat bottom plate) was coated with 100ul/well of the diluted antigen and left overnight at 4°C.

- The following day, the plate was washed 3 times with a washing buffer and dried.
- 100 ul of blocking solution per well was added and incubated at 37°C for 1 hour.
- The plate was washed 3 times and dried.

• The serum samples as well as the negative and positive control sera were diluted 1/100 in a diluting buffer, then 100 ul of each sample was added to two wells from the coated plate and the plate incubated at 37° C for 1 hour.

• The plate was washed 3 times and dried.

• A peroxidase conjugate (anti-species IgG) was diluted 1/800 in the diluting buffer and 50 ul was added to each well, then the plate was incubated at 37°C for 1 hour.

- The plate was washed 3 times and dried.
- 100 ul of the substrate working solution were added to each well.

• The plate was incubated at room temperature in a dark place till a convenient colored reaction develops.

- The reaction was rapidly stopped by addition of 10 ul/well of stopping solution.
- The plate was read on an ELISA reader using a filter of wavelength 490 nm.
- The cut -off point of optical density was 0.1.

The end titre of the tested sample was calculated according to William (1987) as follow:

OD of tested sample - OD of negative control

X titre of positive control

OD of positive control - OD of negative control

3. RESULTS:

3.1.The seroprevalence of EHV-1 specific antibodies in horses and donkeys sera using ELISA in Qalubiah Governorate:

A total of 185 sera samples (77 from horses and 108 from donkeys) were tested for EHV-1 specific antibodies using ELISA. Overall results revealed that 42.86 % (33/77) of the horses sera samples and 37 % (40/108) of the collected donkeys sera samples were EHV-1 seropositive while 39.5% (73/185) of all horses and donkeys sera samples were EHV-1 seropositive. Meanwhile 57.14 % (44/77) in horses and 63 % (68/108) in donkeys sera samples was found to be seronegative for the EHV-1 infections. These results were shown in Tables 3, 4, 5 & Fig. 8.

Table. 3: Seroprevalence of EHV-1 antibodies positive samples in horses and donkeys in	
Qalubiah Governorate.	

Animal	Number of tested	Sero	positive	Seronegative	
	sera	No.	%	No.	0⁄0
Horse	77	33	42.9%	44	57.14%
Donkey	108	40	37%	68	63%
Total	185	73	39.5%	112	60.5%

No. = number of examined animals.

% = Percent.

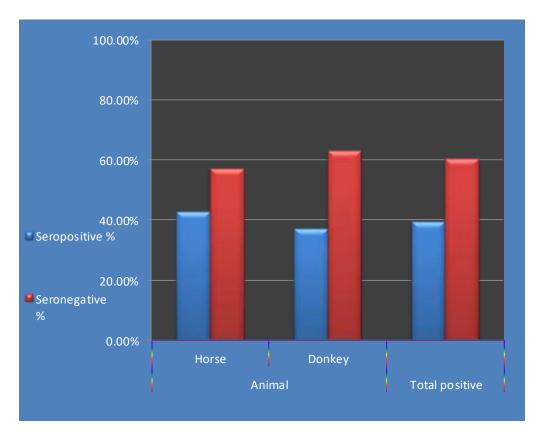


Fig. 8 : Seroprevalence of EHV-1 antibodies positive samples in horses and donkeys in Qalubiah Governorate.

No.	ELISA titer	No.	ELISA titer	No.	ELISA titer	No.	ELISA titer
1	740	24	460	47	490	70	580
2	350	25	69	48	69	71	60
3	90	26	74	49	514	72	566
4	50	27	580	50	64	73	77
5	388	28	66	51	86	74	494
6	46	29	422	52	86	75	499
7	388	30	498	53	70	76	64
8	46	31	59	54	44	77	592
9	80	32	633	55	611		
10	96	33	86	56	65	-	
11	40	34	47	57	590	-	
12	66	35	611	58	74	-	
13	450	36	78	59	48	-	
14	91	37	86	60	671	-	
15	29	38	490	61	55	-	
16	512	39	69	62	680	-	
17	69	40	602	63	593	-	
18	574	41	515	64	69	-	
19	58	42	96	65	530	1	
20	660	43	520	66	77	1	
21	78	44	58	67	82	1	
22	601	45	518	68	429	1	

Table. 4 : EHV-1 antibodies titer of collected horse's sera samples using ELISA in Qalubiah
 Governorate.

23	88	46	58	69	68	
						No. = sample number of

examined horse's sera.

Table. 5 : EHV-1 antibodies titer of collected donkey's sera samples using ELISA in Qalubiah	
Governorate.	

No.	ELISA titer								
1	75	24	528	47	47	70	580	93	520
2	90	25	37	48	610	71	35	94	94
3	515	26	40	49	31	72	598	95	89
4	85	27	522	50	613	73	65	96	579
5	45	28	25	51	87	74	76	97	105
6	590	29	35	52	599	75	603	98	88
7	86	30	710	53	66	76	87	99	518
8	28	31	33	54	535	77	79	100	96
9	602	32	45	55	69	78	480	101	66
10	33	33	634	56	710	79	98	102	599
11	40	34	96	57	87	80	83	103	87
12	509	35	88	58	93	81	595	104	92
13	35	36	598	59	528	82	71	105	520
14	27	37	84	60	65	83	87	106	97
15	701	38	87	61	720	84	564	107	498
16	44	39	589	62	28	85	98	108	88
17	38	40	89	63	548	86	93		
18	528	41	99	64	85	87	604	1	
19	42	42	558	65	498	88	86	-	

20	605	43	76	66	90	89	89	No. = sample
21	37	44	559	67	538	90	79	number of
22	599	45	36	68	87	91	585	examined donkey's sera.
23	32	46	40	69	79	92	59	j

3.2. Age, sex and species susceptibility:

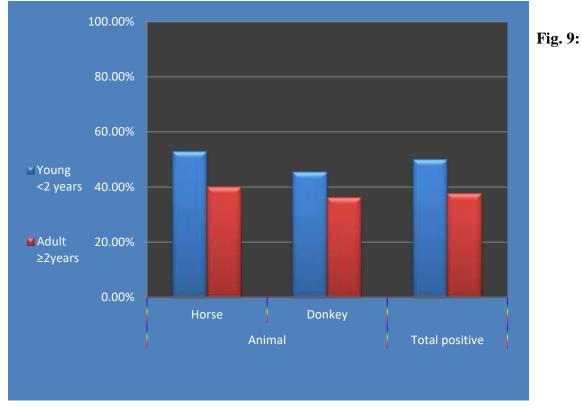
Among (185) horses and donkeys sera screened by ELISA, (73) were found positive for EHV-1 antibodies. Out of (73) positive cases, (33) were found positive for horses sera and (40) were found positive for donkeys sera.

As shown in Table. 6 and Fig. 9. Analysis of data revealed that, young animals (<2 years) were represent (50%) (14/28) while adult animals (\geq 2years) were (37.6%) (59/157) EHV-1 antibodies seropositive. Further analysis showed that, the EHV-1 antibodies seropositive young horses (9/17) (52.9%) were higher than their adults (40%) (24/60). While EHV-1 antibodies seropositive young donkeys (45.5%) (5/11) were higher than their adults (36.1%) (35/97). Table. 6: Seroprevalence of positive EHV-1 antibodies in sera of young and adult horses and donkeys using ELISA in Qalubiah Governorate.

Species		An	Total positive		
Age		Horse	Donkey		
Young <2 years	No.	9	5	14	
- J U	%	52.9% (9/17)	45.5% (5/11)	50% (14/28)	
Adult ≥2years	No.	24	35	59	
_ ,	%	40% (24/60)	36.1% (35/97)	37.6% (59/157)	
Total		33	40	73	

No. = number of tested animals.

% = Percent of positive sample from the total number of the same age group.



Seroprevalence of positive EHV-1 antibodies in sera of young and adult horses and donkeys using ELISA in Qalubiah Governorate.

In the present study, the total number of (185) examined horses and donkeys sera were differentiated into (79) males and (106) females. As shown in Table. 7 and Fig. 10. Out of (79) male cases, 15.2% (12/79) of total male horses and donkeys were EHV-1 seropositive while 15.15% (5/33) of male horses sera and 15.2% (7/46) of male donkeys sera were EHV-1 seropositive. However, Out of (106) female cases, 57.5% (61/106) of total female horses and donkeys were EHV-1 seropositive while the percentage of seropositive sera samples of female horses and donkeys were 63.6% (28/44) and 53.2% (33/62) respectively.

Species		Animals		Total positive
		Horses	Donkeys	
Sex				
Male	No.	5	7	12
	%	15.15% (5/33)	15.2% (7/46)	15.2% (12/79)
Female	No.	28	33	61
	%	63.6% (28/44)	53.2% (33/62)	57.5% (61/106)
Total	No.	33	40	73
	%	42.9% (33/77)	37% (40/108)	39.5% (73/185)

Table. 7 : Seroprevalence of EHV-1 antibodies positive sera of male and female horses and donkeys using ELISA in Qalubiah Governorate.

number of tested animals.

% = Percent of positive sample from the total number of the same sex group.

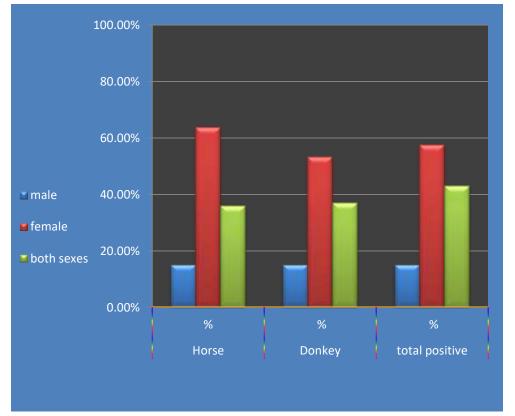


Fig. 10 : Seroprevalence of positive EHV-1 antibodies in sera of male and female horses and donkeys using ELISA in Qalubiah Governorate.

4. DISCUSSION:

In Egypt, the first recorded result for the presence of EHV-1 specific antibodies in horse's serum through serological evidence was done by (**Matumoto et al., 1965**), while EHV-1 was first isolated by (**Hassanein et al., 2002**) in Cairo from aborted mares at the last trimester of pregnancy.

The serological investigation of EHV-1 that conducted in this study aimed to determine the prevalence of the disease, as well as the age and sex susceptibility.

Regarding the seroprevalence of EHV-1, out of 185 sera sample (77 from horses and 108 from donkeys) examined for EHV-1 specific antibodies using ELISA. Overall results revealed that 42.86 % (33/77) of the horses sera samples and 37 % (40/108) of the collected donkeys sera samples were EHV-1 seropositive while 39.5% (73/185) of total horses and donkeys sera samples were EHV-1 seropositive as shown in **Tables (3, 4, 5)**. The same results obtained by (**Avci, et al., 2014**) who recorded that, 34.66% (52/150) were found to be positive for EHV-1 antibodies. Also

this result agrees with (**Abdelgawad, et al., 2015**) who reported, that 33.3% (4/12) EHV-1 seropositive in donkey by using peptide-based ELISA. (**Ataseven** *et al.*, **2009**) revealed that the seropositivity for EHV-1 was 14.5%. However, other studies, reported higher levels of infection; for example: (**Yildirim et al., 2015**) who reported that 52.48% (222/423) of the horses sampled and 51.85% (126/243) of the donkeys sampled were EHV-1 seropositive. When the horse and donkey samples were evaluated together, 52.25% were seropositive for EHV-1. Also (**Wegdan, et al., 2016**) reported that 65.9% of the total horse and donkeys serum samples examined were seropositive for EHV-1.

None of the animals that tested positive for antibodies to EHV-1 exhibited clinical symptoms indicative of herpesvirus infection. This is characteristic of natural viral host species (**Abdelgawad** et al., 2015).

Regarding the Age, sex and species susceptibility, analysis of data revealed that, young animals (<2 years) were represent (50%) (14/28) while adult animals (\geq 2 years) were (37.6%) (59/157) EHV-1 antibodies seropositive. Further analysis showed that, the EHV-1 antibodies seropositive young horses (52.9%) (9/17) were higher than their adults (40%) (24/60). While EHV-1 antibodies seropositive young donkeys (45.5%) (5/11) were higher than their adults (36.1%)(35/97). As shown in Table (6) & Fig. (9). The same results were obtained by (Pusterla, et al., 2016) who reported that EHV-1 generally affects horses less than three years of age with the majority of affected horses being less than one year of age. Although study cases represented a wide range of ages, 65% of them were ≤ 10 years of age. This reflects the higher susceptibility of young animals to infectious respiratory pathogens (Mumford and others 2003). Also (Foot et al., 2004) reported that, infections caused by EHV-1 are particularly common in young performance horses. (Allen, 2008) estimated that 80 to 90% of horses have been exposed to EHV1 infections by two years of age and respiratory disease associated with EHV1 is most commonly seen in young animals at the time of weaning. While experimental infection proved that older horses are more predisposed to the development of neurological disease as compared to young to young/middle aged horses. Adult horses may develop viremia 100 times higher than young horses and they are 8 times more likely to develop the disease.

In the present study, the total number of (185) examined horses and donkeys sera were differentiated into (79) males and (106) females. Out of (79) male cases, 15.2% (12/79) of total male horses and donkeys were EHV-1 seropositive while 15.15% (5/33) of male horses sera and

15.2% (7/46) of male donkeys sera were EHV-1 seropositive. However, Out of (106) female cases, 57.5% (61/106) of total female horses and donkeys were EHV-1 seropositive while 63.6% (28/44) of female horses sera and 53.2% (33/62) of female donkeys sera were EHV-1 seropositive. As shown in **Table (7) & Fig. (10).** This result came in similarity with results of (**Goehring, et al., 2006**) who reported higher infection rate among mares in Netherlands. This higher detection among females differ with the findings of (**Hafshejani et al., 2015**) in Isfahan who suggest that the stallions EHV-1: 18.18% were infected more than the mares EHV-1: 9.67%, and therefore had more active infection by the viruses. But the result of Shahrekord (EHV-1: stallion (8.96%), mare (7.14); suggested otherwise. This finding suggests that sex may not have played a role in EHV infection. (**Momtaz and Hematzadeh 2003**) reported that sex did not significantly affect infection of horses in Chaharmahal and Bakhtiari province, Iran. Other authors elsewhere had reported that sex is a factor in the epidemiology of infection by EHV-1 (**Goehring, et al., 2006 and Lunn, et al., 2009**). The differences in infection rate among sexes in these studies may be due to differences in rate of exposure to infection, health status (such as pregnancy and suckling in mares), age, previous vaccinations, or immune status of the animals sampled in the study areas.

5. References:

- Abdelgawad, A., Hermes, R., Damiani, A., Lamglait, B., Gábor Á. Czirják, East, M., Aschenborn, O., Wenker, C., Kasem, S., Osterrieder, N., Greenwood, A.D. (2015): Comprehensive Serology Based on a Peptide ELISA to Assess the Prevalence of Closely Related Equine Herpesviruses in Zoo and Wild Animals PLOS ONE | DOI:10.1371/journal.pone.0138370 September 17.
- Allen, G.P., (2008): Risk factors for development of neurologic disease after experimental exposure to equine herpesvirus-1 in horses. Am. J. Vet. Res. 69,1595–1600.
- American Association of Equine Practitioners, AAEP (2013): Equineherpesvirus-1 and4 related diseases.
- Ataseven, V.S., Dagalp, S.B., Guzel, M., Basaran, Z., Tan, M.T., Geraghty, B., (2009): Prevalence of equine herpesvirus-1 and equine herpesvirus-4 infections in equidae species in Turkey as determined by ELISA and multiplex nested PCR. Res. Vet. Sci. 86, 339–344.

- Avci, O., Yavru, S., Tokgoz, S. and Kale, M. (2014): Detection of Antibodies against Equine Herpes Virus-1 and Equine Herpes Virus-4 in Horses in Southeast Anatolia by Indirect ELISA; Acta Scientiae Veterinariae, 42: 1250.
- Azmi, M. & Field, H. J. (1993): Interactions between equine herpesvirus type 1 and equine herpesvirus type 4: T cell responses in a murine infection model. Journal of general virology2345-2339,74,
- Dutta, S. K.; Talbot, N. C. and Myrup, A. C. (1983): Detection of Equine herpesvirus-1 antigen and the specific antibody by enzyme linked immunosorbent assay. Am. J. Vet. Res.; 44 (10): 1930-1934.
- Foote, C.E., Love, D.N., Gilkerson, J.R., Whalley, J.M. (2004): Detection of EHV-1 and EHV-4 DNA in unweaned Thoroughbred foals from vaccinated mares on a large stud farm. Equine Vet J.;36:341–5.
- Gilkerson, JR, Teague, N, Whalley, JM, Love, DN. A prospective cohort study of upper respiratory tract disease in one and two year old racehorses. Serological evaluation of the role of equine herpesviruses 1 and 4 (EHV-1 and EHV-4) in respiratory disease. Aust Equine Vet. 1999;17:76–81.
- Goehring, L.S., van Winden, S.C., van Maanen, C., Sloet van, Oldruitenborgh-Oosterbaan, M.M. (2006): Equine herpesvirus type 1-associated myeloencephalopathy in The Netherlands: a four-year retrospective study (1999 –2003). J. Vet. Intern. Med. 20:601–607.
- Hafshejani, T.T., Nekoei, S., Vazirian, B., Doosti, A., Khamesipour, F. and Anyanwu, M.U.
 (2015): Molecular Detection of Equine Herpesvirus Types 1 and 4 Infection in Healthy Horses in Isfahan Central and Shahrekord Southwest Regions, Iran. Hindawi Publishing Corporation BioMed Research International Volume 2015, Article ID 917854, 7 pages <u>http://dx.doi.org/10.1155/2015/917854</u>
- Hassanein, M. M.; Maysa, H.; El-Bagoury, F.; Magda, A. K.; EL-Kabbany, M. M. A. and Daoud, M. A. (2002). Trials for isolation and identification of equine herpesvirus abortion in Egypt. Vet. Med. J., Giza, Vol.; 50(4): 977-986.
- Lunn D.P., Davis-Pointer N., Flaminio M.J., Horohov D.W., Osterrieder K., Pusterla N. & Townsend H.G. 2009. Equine herpesvirus-1 consensus statement. *Journal of Veterinary Internal Medicine*. 23(3): 450-461.



- Matumoto, M.; Ishizaki, R. and Shimizu, T. (1965): Serological survey of equine rhinophneumonitis virus infection among horses in various countries. Arch. Ges. Virusforsh; 15: 509-624.
- Momtaz, H. and Hematzadeh, F. (2003): "ASerological survey on equine herpes virus 1 and equine herpes virus 4 in the horse using

ELISA," Pajouhesh & Sazandegi, vol. 59, pp. 63-69,

- Mumford, E.L., Traub-Dargatz, J.L., Carman, J., Callan, R.J., Collins, J.K., Goltz, K.L., Romm, S.R., Tarr, S.F. and Salman, M.D. (2003) Occurrence of Infectious Upper Respiratory Tract Disease and Response to Vaccination in Horses on Six Sentinel Premises in Northern Colorado. Equine Veterinary Journal, 35, 72-77.
- **OIE**, (2015): manual of diagnostic tests and vaccines for: Equine rhinopnumonitis (Equine Herpes virus 1 and 4), chapter 2.5.9.

OIE (2015). Equine rhinopenumonitis. Manual of standards diagnostic test and vaccines. 5th Ed. Par 2, Chapter 2.5.9:1-12.

- Patel, J.R. and Heldens, J. (2005): Equine herpesviruses 1(EHV-1) and 4(EHV-4) epidemiology, disease and immunoprophylaxis: a brief review .The Veterinary Journal, 170, 14–23.
- Pusterla, N., Mapes, S., Akana, N., Barnett, C., Mackenzie, C., Gaughan, E., Craig, B., Chappell, D. & Vaala, W. (2016): Prevalence factors associated with equine herpesvirus type 1 infection in equids with upper respiratory tract infection and/or acute onset of neurological signs from 2008 to 2014. Veterinary Record, 178, 70-70.
- Siedek M.E., Whelan M., Edington N. & Hamblin A. (1999). Equine herpes virus type 1 infects dendritic cells in vitro: Stimulation of T lymphocyte proliferation and cytotoxicity by infected dendritic cells. Veterinary Immunology and Immunopathology. 67(1): 17-32.

Van Maanen, C. (2002): Equine herpes virus 1 &4 infections; an update. Vet.Quarterly24:58-78.

Wegdan, H.A., Intisar K. S., Shaza, M. M., Algezoli O. A., Ballal A., Ihsan H. A., Sahar M.
E., Baraa A. M., Manal H. S., Muna E. A., Taha K. M., Nada E. M. and Ali Y. H.
(2016): Serological Detection of Equine Herpes Virus (EHV) Type 1 and 4 in Sudan British Microbiology Research Journal 14(6): 1-6, Article no.BMRJ.25803

Williams, R. (1987): ELISA technique for diagnosis of AHSV. J. Vet. Diag. Invest., 11 (2): 9-11.

- Yasunaga S., Maeda K., Matsumura T., Kai K., Iwata H. & Inoue T. (1998): Diagnosis and sero-epizootiology of equine herpesvirus type 1 and type 4 infections in Japan using a type-specific ELISA. The Journal of Veterinary Medicine Science. 60(10): 1133-1137.
- Yildirim, Y.; Yilmaz, V. and Kirmizigul, A. H. (2015): Equine herpes virus type 1 (EHV-1) and 4 (EHV-4) infections in horses and donkeys in northeastern Turkey Iranian Journal of Veterinary Research, Shiraz University, Vol. 16, No. 4, Ser. No. 53, Pages 341-344